

**A hierarchical map of orthologous genomic regions reconstructed from two very closely related genomes: A case study of the cucumber (*Cucumis sativus*)**

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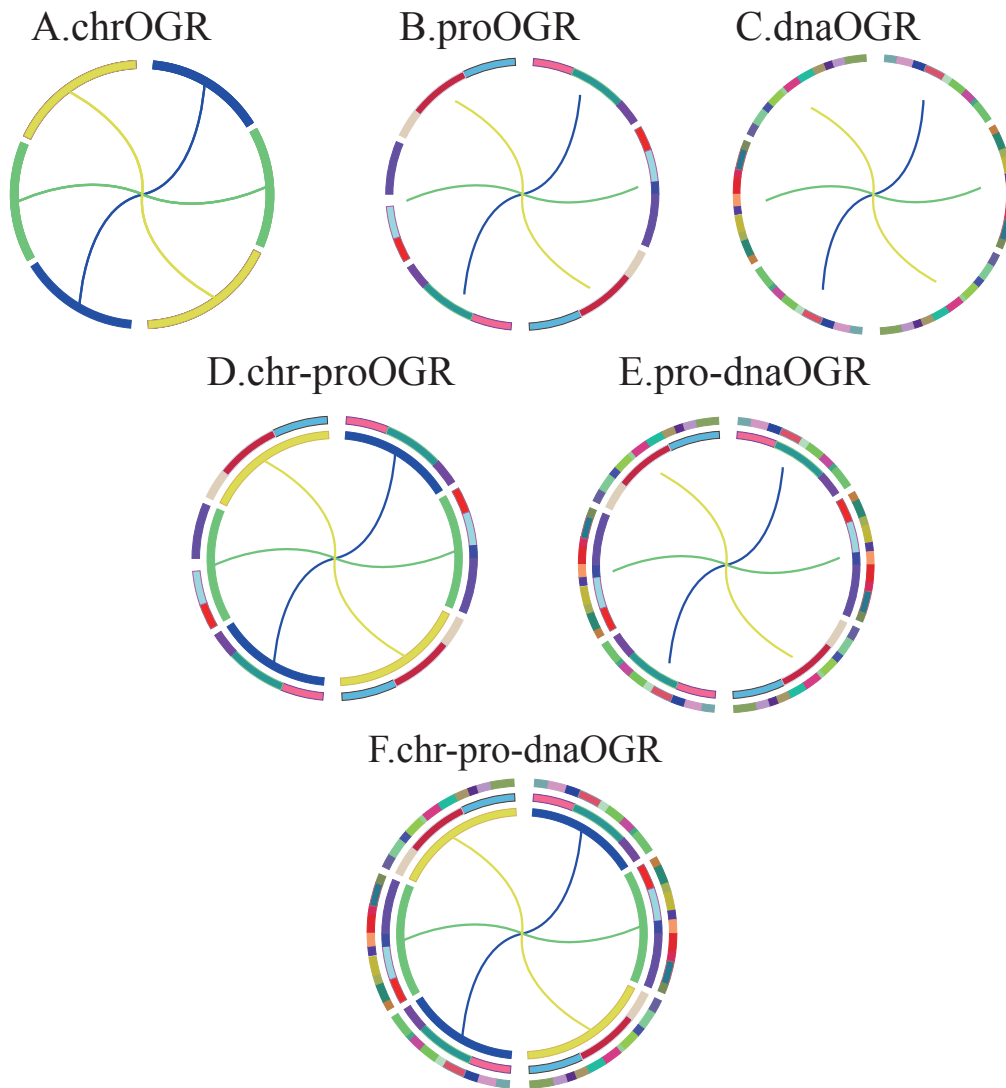
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Life Sciences, Beijing Normal University, Beijing 100875, China.

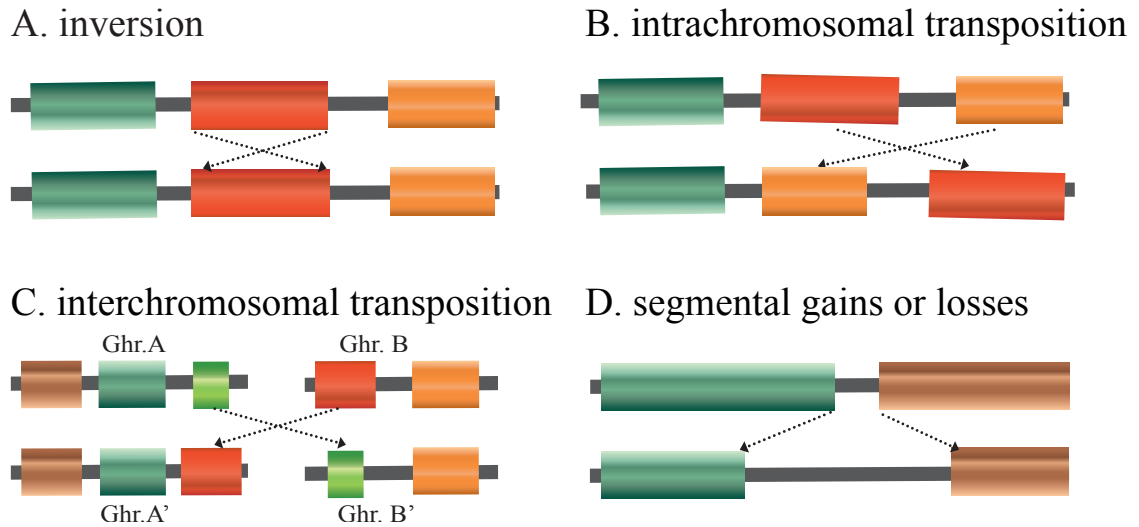
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**Supplementary Material**

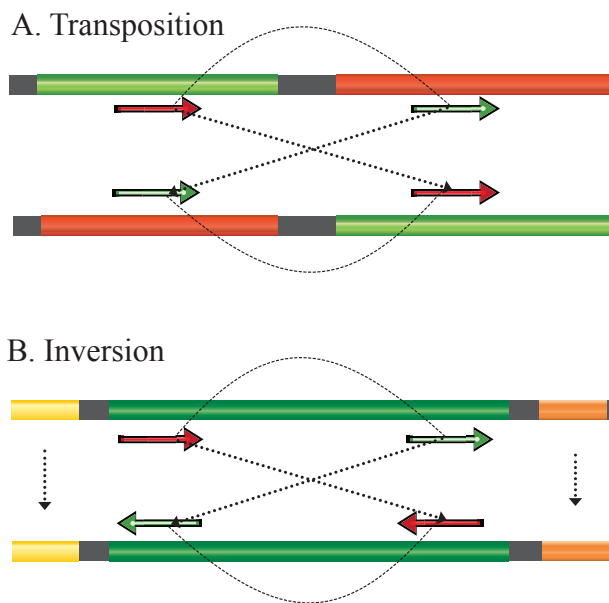
## Supplementary Figures:



**Figure S1. The schematic diagram of constructing hierarchical map of OGRs between closely related genomes.** The OGRs from different levels were based on different types of markers. Each high-level OGR would contain one or more low-level OGRs. Based on the observation, we constructed a hierarchical map of OGRs. (A) The chrOGRs were detected by the previous work (SSRs as markers). (B) The proOGRs were detected by i-ADHoRe (protein-coding genes as markers). (C) The dnaOGRs were detected by Mugsy (conserved DNA sequences as markers). (D) The chr-proOGRs were constructed by mapping the proOGRs onto the chrOGRs. (E) The pro-dnaOGRs were constructed by mapping the dnaOGRs onto the proOGRs. (F) The three-level OGRs (chrOGRs:proOGRs:dnaOGRs) were constructed by integrating the chr-proOGRs and pro-dnaOGRs. Finally the three-level hierarchical map of OGRs were constructed.



**Figure S2. Configurations of large-scale genomic changes detected in the hierarchical map of OGRs.** The large-scale genomic changes were detected by comparing the structural difference between its respective chromosomal segments from the two genomes, while treating their respective child OGRs as invariant markers. (A) When the orientations of two segments of the child OGR were opposite, this child OGR would be assigned as an inversion candidate. (B) If the orders of the child OGRs between its respective chromosomal segments from the parent OGR were different and the two segments of the child and parent were from the same chromosome, respectively, this child OGR would be treated as an intrachromosomal transposition. (C) If one chromosome from the child OGR was different with the chromosome of the parent OGR, this child OGR would be assigned as interchromosomal transposition. (D) If the regions of the parent OGR were not overlapped with the child OGRs, these regions would be treated as segmental gains or losses. The colorful bar represents the regions located at OGRs and the grey bar represents the regions not located at OGRs.



**Figure S3. Configurations of verified large-scale genomic changes by pair-end reads.**

We used the pair-end reads to verify detected large-scale genomic changes: (a) Transposition, where the orders of two pairs of reads are different but the orientations are same; (b) inversion, where the orders from two pairs of reads are different and the orientations are opposite. The colorful bar represents the regions are located at OGRs and the grey bar represents the regions that are not located at OGRs.

## Supplementary Tables:

**Table S1. The websites and versions of the five genomes**

Genomes	Version	The Date of Download	Websites
<i>Cucumis sativus L.</i>	V2	2014/08/10	<a href="http://cmb.bnu.edu.cn/Cucumis_sativus_v20/">http://cmb.bnu.edu.cn/Cucumis_sativus_v20/</a>
<i>C. sativus var. hardwickii</i>	V1	2014/08/10	<a href="http://cmb.bnu.edu.cn/Cucumis_sativus_v20/resequencing/">http://cmb.bnu.edu.cn/Cucumis_sativus_v20/resequencing/</a>
<i>Cucumis melo L.</i>	V3.5	2012/10/17	<a href="https://melonomics.net/files/Genome/">https://melonomics.net/files/Genome/</a>
<i>Citrullus lanatus</i>	V1	2013/09/21	<a href="ftp://www.icugi.org/pub/genome/watermelon/">ftp://www.icugi.org/pub/genome/watermelon/</a>
<i>Solanum lycopersicum</i>	V2.3	2013/08/08	<a href="ftp://ftp.solgenomics.net/tomato_genome/annotation/ITAG2.3_release/">ftp://ftp.solgenomics.net/tomato_genome/annotation/ITAG2.3_release/</a>

**Table S2. Numbers of gene families (and genes) in five plant genomes**

Genomes	Singletons <sup>†</sup>	Two-genes <sup>‡</sup>	$\geq 3$ <sup>§</sup>	Total gene families <sup>¶</sup>
<i>Cucumis sativus L.</i>	16,948(16,948)	1,032(2,064)	435(2,008)	18,415(21,020)
<i>C. sativus var. hardwickii</i>	16,840(16,840)	1,018(2,036)	507(2,724)	18,365(21,600)
<i>Cucumis melo L.</i>	14,703(14,703)	1,291(2,582)	795(5,248)	16,789(22,533)
<i>Citrullus lanatus</i>	14,221(14,221)	1,109(2,582)	652(4,446)	15,982(20,885)
<i>Solanum lycopersicum</i>	9,065(9,065)	3,076(6,152)	1,675(10,150)	13,816(25,367)
All five genomes	71,777(71,777)	7,526(15,052)	4,064(24,576)	21,488(111,405)

<sup>†</sup>The number of singletons.

<sup>‡</sup>The number of gene families (genes) with two genes.

<sup>§</sup>The number of gene families (genes) contain at least three genes.

<sup>¶</sup>The number of total gene families (genes).

**Table S3. The statistics of dnaOGRs overlapped with proOGRs (in supported excel)****Table S4. The statistics of dnaOGRs overlapped with chrOGRs (in supported excel)****Table S5. The statistics of length distributions of large-scale genomic changes at proOGR level**

Type	No. of genomic changes	Min. of length	Max. of length	Mean of length	Total length
Intrachromosomal					
transposition	386	30	316,528	2,997	1,156,715
Interchromosomal					
transposition	5,304	30	12,826	240	1,273,820
Inversion	2,480	30	73,631	675	1,673,537
Segmental gain or loss	5,939	30	15,567	643	3,820,578

**Table S6. The distribution of the SNVs and Indels at dnaOGR level**

Feature	No. of SNVs	No. of indels	No. of kept SNVs	No. of kept indels	Ratio of kept SNVs	Ratio of kept indels
Gene	395,546	81,716	222,682	43,429	36.73%	27.32%
Exon	108,837	13,464	70,859	7,862	11.69%	4.95%
CDS	79,077	4,159	52,717	2,098	8.70%	1.32%
5'utr	9,738	3,107	6,201	2,021	1.02%	1.27%
3'utr	20,022	6,198	11,941	3,743	1.97%	2.35%
Intron	275,312	65,056	145,449	33,752	23.99%	21.24%
Intergenic	1,628,026	399,422	383,373	115,450	63.23%	72.64%
ncRNA	1,162	362	214	62	0.04%	0.04%
Total	2,024,734	481,500	606,269	158,941	100.00%	100.00%

**Table S7. The statistics of the length distributions of large-scale genomic changes at chrOGR and dnaOGR levels**

Type	No. of genomic changes	Min. of length	Max. of length	Mean of length	Total length
Intrachromosomal transposition <sup>†</sup>	14	28,752	167,847	77,386	1,083,399
Interchromosomal transposition <sup>†</sup>	3	29,689	53,882	42,447	127,340
Segmental gain or loss <sup>†</sup>	542	666	493,561	42,190	22,867,006
Segmental gain or loss <sup>‡</sup>	38,620	30	2,279	170	6,551,635

<sup>†</sup> Genomic changes at chrOGR level

<sup>‡</sup> Genomic changes at dnaOGR level

**Table S8. The number of synonymous and nonsynonymous SNVs**

Type	Synonymous	Nonsynonymous	Stop-codon SNVs
SNVs	12,163	66,914	3,082
Kept SNVs	8,559	44,158	2,117

**Table S9. The statistics of length distributions of verified large-scale genomic changes at proOGR level**

Type	No. of genomic changes	Min. of length	Max. of length	Mean of length	Total length
Intrachromosomal					
transposition	23	70	316,528	19,686	452,789
Interchromosomal					
transposition	23	70	316,528	19,686	452,789
Inversion	174	32	73,631	5,736	998,150
Segmental gain or loss	1,726	30	15,567	1,129	1,948,044

**Table S10. The comparison of previous detected genomic changes and our detected genomic changes**

Type	No. of Genomic Changes	No. of Overlapped Genomic Changes	Ratio of Overlapped Genomic Changes
Segmental gains or losses <sup>†</sup>	1,936	1,922	99%
Inversions <sup>†</sup>	7	5	71%
SNVs <sup>‡</sup>	1,421,881	1,220,768	86%
Small Indels <sup>‡</sup>	18,420	2,814	15%

<sup>†</sup>The large-scale genomic changes were from the work of Zhang (Zhang, Mao, et al., 2015).

<sup>‡</sup>The small-scale genomic changes were unpublished and from the work of Qi (Qi, Liu, et al., 2013).

**Table S11. The number of frameshift indels**

Type	Frameshift	Non-frameshift
Indels	2,330	1,829
Kept indels	693	534

**Table S12. Statistics of overlapped proportions of detected SNVs and small indels at dnaOGR level with results detected by reads mapping**

	No. of SNVs	No. of Indels	No. of shared SNVs	No. of shared indels	Ratio of SNVs overlap	Ratio of indel overlap
Raw genomic changes	2,024,734	481,500	1,623,617	138,709	80%	29%
Kept genomic changes	606,269	158,941	575,779	64,760	95%	41%

**Table S13. The top 20 GO terms of verified large-scale genomic changes (in supported excel)**

**Table S14. The top 20 GO terms of verified small-scale genomic changes (in supported excel)**

**Table S15. The coordinates in the two cucumber draft genome assemblies of the chrOGRs (in supported excel)**

**Table S16. The coordinates in the two cucumber draft genome assemblies of the proOGRs (in supported excel)**

**Table S17. The coordinates in the two cucumber draft genome assemblies of the dnaOGRs (in supported excel).**

Qi, J., X. Liu, D. Shen, H. Miao, B. Xie, X. Li, et al. 2013. A genomic variation map provides insights into the genetic basis of cucumber domestication and diversity. *Nature Genet.* 45: 1510-U1149. doi:10.1038/ng.2801.

Zhang, Z., L. Mao, H. Chen, F. Bu, G. Li, J. Sun, et al. 2015. Genome-Wide Mapping of Structural Variations Reveals a Copy Number Variant That Determines Reproductive Morphology in Cucumber. *Plant Cell* 27: 1595-1604. doi:10.1105/tpc.114.135848.